EFFICACY OF SACCHAROMYCES BOULARDII AS A PROBIOTIC IN DOGS WITH LINCOMYCIN INDUCED DIARRHOEA

MUSTAFA SINAN AKTAS, MEHMET KAZIM BORKU¹, AND YUNUSEMRE OZKANLAR

Department of Internal Medicine, College of Veterinary Medicine,
Ataturk University, 25700, Erzurum, Turkey

¹Department of Internal Medicine, College of Veterinary Medicine,
Ankara University, 06110 Ankara, Turkey
sinanaktas@atauni.edu.tr

Received for publication February 27, 2007

Abstract

Twenty-four dogs were divided into three equal groups. Lincomycin was administered intramuscularly to all groups. However, group II was given S. boulardii after lincomycin induced diarrhoea occurred, and in group III, lincomycin was administered together with S. boulardii. Lincomycin caused diarrhoea in groups I (in 7.8±1.6 d) and II (in 6.9±2 d), but it did not cause diarrhoea in group III. The durations of diarrhoea were significantly different (P<0.05) between group I (6.5 \pm 1.2 d) and group II (2.9 \pm 0.4 d). The total short chain fatty acid (TSCFA) level was significantly lower (P<0.001) compared with their baselines in groups I and II at the time of the diarrhoeas' onset and cessation. Additionally, there were differences between groups at the time of diarrhoea cessation and 1 week after the treatment (P<0.05). In conclusion, lincomycin decreased TSCFAs causing diarrhoea in the dogs when given alone, and S. boulardii was effective in treating lincomycin induced diarrhoea and to prevent the occurrence of diarrhoea when given together with lincomycin.

Key words: dog, diarrhoea, *Saccharomyces boulardii*, lincomycin, short chain fatty acids.

Antibiotics may cause side effects as seen with other drugs (14). Diarrhoea as a complication is observed in 5%-25% of humans who take antibiotics (3). Ampicillin, lincomycin, clindamycin, and cefalosporins were reported as a of cause diarrhoea (27). Some antibiotics and chemicals cause the deterioration in normal intestinal flora and result in the growth of some pathogenic microorganisms (4, 24). Clostridium difficile (Cl difficile), growing faster than other pathogens existing in the colon, has been the most frequently seen pathogen in antibiotic induced diarrhoea (24). Besides, it was noted that the other pathogens (3%) cause diarrhoea as well (23).

In vitro studies showed that the short chain fatty acid (SCFA) values decreased, when most of the

antibiotics reach sufficient concentration levels in the colon (19). By decreasing the SCFA content, absorption of water and electrolytes decrease and diarrhoea occurs. When the level of carbohydrates that cannot be absorbed in the intestine lumen increases, they take water by an osmotic effect and cause diarrhoea (38).

Saccharomyces boulardii (S. boulardii), a member of Saccharomycetaceae family, is yeast that has an ability of suppression and fermentation of the carbohydrates showing Gram+ staining property. As yeast, it is resistant to antibiotics showing a difference from probiotics of a bacterial origin. It is not affected by gastric acid and bile secretion (12). S. boulardii competes for food and mucosal receptors with pathogenic microorganisms in the intestine lumen (15), and destroys bacterial toxins and their receptor sides by releasing proteases (11).

In humans, one of the treatments of choice is *S. boulardii* as a probiotic for the therapy of antibiotic induced diarrhoea (5, 37). An increase in aminopeptidase and disaccharidase levels was detected in rats given *S. boulardii* (9). However, to our knowledge there is no report on the use of *S. boulardii* as a probiotic in such cases in dogs. The purpose of this study was to investigate the efficacy of *S. boulardii* in dogs with lincomycin induced diarrhoea.

Material and Methods

The Ethic Committee of Animal Care and Use in the Faculty of Veterinary Medicine in Ankara University approved this experimental protocol (Decision No. 2004/11). Different breeds of dogs, 14 male and 10 female, aging 1-5 years were used in the study. Vaccination and anti-parasite drug administration were applied to all dogs. The dogs were fed a commercial dog feed and had water *ad libitum*.

The dogs were divided into three equal groups. Dogs in the groups I and II were injected intramuscularly (i.m.) with 150 mg/kg/d of lincomycin (Linkoles, Aroma, Turkey) until diarrhoea occurred. After the onset of diarrhoea, group II was given 1000 mg/d of *S. boulardii* (Reflor, Dif, Turkey) orally for ten days. Group III was given i.m. for ten days 150 mg/kg/d of lincomycin together with an oral dose of 1000 mg/d of *S. boulardii*.

Sampling and clinical examinations in groups I and II were carried out four times, i.e. before the experiment (baseline), just after the diarrhoea occurred (onset of diarrhoea) and stopped (diarrhoea cessation), and a week after the diarrhoea stopped (1 week after treatment). Sample collection and examinations in group III was performed twice, i.e. before the experiment (baseline) and on the 10th d of the study. Clinical findings included respiration and pulse rates, body temperature, appearance, and the existence of diarrhoea, dehydration, appetite, and behaviour. Haemogram, SCFAs, and Cl. difficile toxins (Ridascreen Cl. difficile toxin-A/B EIA) were analysed in faeces on the related days. The isolations of pathogenic microorganisms such as E. coli, Shigella, Salmonella, and Campylobacter were performed by culturing the faeces. The faecal smears were examined for possible spores of Cl. perfiringens as well (25).

The analyses of total short chain fatty acids (TSCFAs) were carried out using gas chromatography (Agilent Technologies 6890 Network GC system). A sample of 2-5 g faeces was homogenised with 4 ml of 2-ethylbutiric acid (3 mmol/L) and 1 ml of $\rm H_2SO_4$ (0.5 mmol/L), and then the vacuum distillation was performed. Two micro litre samples were used by an automatic integrator to obtain the peak values of each short chain fatty acid. Comparing the peak values of standards and samples each to other, TSCFAs were calculated in mmol/L units (21). TSCFAs included all of the SCFA measurements.

Statistical analysis was done using a One-Way and Two-Way ANOVA test, incorporating a repeated measures design and if indicated by a significant F-statistic, differences in specific means were sought by a Tukey's post hoc test requiring a P<0.05 for significance.

Results

Six out of 8 dogs (75%) in the group I, 7 (87.5%) in group II, and none in group III, developed diarrhoea. The time for the onset of diarrhoea after lincomycin administration and duration of diarrhoea for groups I and II, are shown in Table 1. The time for the onset of diarrhoea in both groups was noted as similar. The duration of diarrhoea in group II was significantly (P<0.05) shorter than that of group I, showing the effect of the treatment.

Table 1
Mean days for the onset of diarrhoea after lincomycin administration and duration of diarrhoea in groups I and II

	Time	Group I	Group II
	Onset of diarrhoea	7.8 ± 1.6	6.9 ± 2.0
	Duration of diarrhoea	6.5 ± 1.2	2.9 ± 0.4
±: standard deviations of the means			

In the physical and haematological examination of the groups I and II, no abnormalities were detected, except for diarrhoea; and any abnormalities (data are not shown) were displayed in group III. No *Cl. difficile* toxin was found, and no pathogen isolation took place in any of the groups throughout the experiment.

The mean values of TSCFAs in groups I and II are shown as the time-points of baseline, onset, and cessation of diarrhoea, and 1 week after the treatment (Fig. 1). The mean values of TSCFAs in both groups decreased significantly (P<0.001) at the time of the onset of diarrhoea and slightly increased with the same manner during the course of the experiment for up to 1 week after the treatment. However, TSCFA levels in the group I was lower than its baseline 1 week after the treatment, while that in the group II reached the steady position of its baseline (P=0.127). When comparing both groups, the values of TSCFAs were higher in group II than in group I at the time-points of diarrhoea cessation and 1 week after the treatment (P<0.05).

The mean values of TSCFAs in group III were demonstrated for the time-points before the experiment (baseline) and on the 10th d of its course (Fig. 2). There were no statistically significant differences between the time-points of data collection (P=0.1).

Discussion

Berezin (4) reported that diarrhoea might usually develop in 3-9 d after commencement of the administration of antibiotics. Most of the dogs in the presented study exhibited diarrhoea in the first and second group as a side effect of lincomycin. A significant difference (P<0.05) occurred between both groups concerning the duration of diarrhoea, suggesting that *S. boulardii* is an effective cure for the induced diarrhoea.

Various antibiotics given to hamsters (6), pigs (36), bears (30), penguins (18), mares and their offspring, adult horses, rats (2), rabbits (33), and stray dogs (28) cause diarrhoea, and in some cases, *Cl. difficile* toxins have been isolated as an active agent. Some researchers (7, 16, 34, 40) claimed that *Cl. difficile* toxins were rarely encountered in diarrhoea caused by antibiotics in dogs. In this study, *Cl. difficile* toxins were not isolated in the cases of diarrhoea caused by lincomycin. Boon and Beale (6) indicated that a dose related *C. difficile* toxin appeared in a hamster.

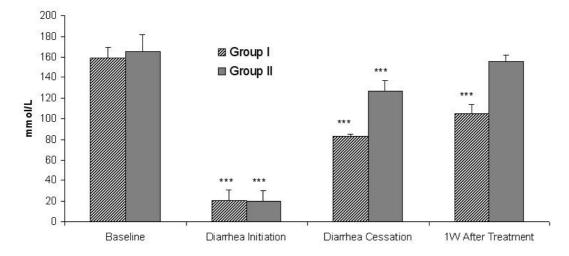


Fig. 1. Mean values of total short chain fatty acids of baseline, diarrhoea initiation, diarrhoea cessation, and 1 week after treatment. *** P<0.001 compared with the baseline in each group. The bars represent standard deviations of the means.

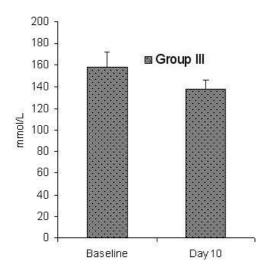


Fig. 2. Mean values of total short chain fatty acids of baseline and 10th d of the experiment. The bars represent standard deviations of the means.

It has also been mentioned that, apart from Cl. difficile, some other pathogenic organisms could cause diarrhoea by multiplying (3%) themselves in the intestines. E. coli, Salmonella, Campylobacter, Cl. perfiringens, and Shigella sp. have been considered as enteric pathogenic microorganisms that could cause diarrhoea in dogs (26). The fact that more than five Cl. perfiringens spores were found in a microscopic immersion field of faecal smears obtained from the dogs with diarrhoea; suggests that diarrhoea may be associated with Cl. perfiringens (39). Any faecal pathogenic microorganism did not multiplied in the cultures and no Cl. perfiringens spores was detected in any of the faecal smears in the dogs given lincomycin.

Further studies in dogs should be addressed to highlight the effects of a longer duration and/or a higher dosage of lincomycin than the experimental design was used here, in order to reveal the potential existence of pathogenic organisms given above.

Acute infections, foreign proteins, tissue damage, blood loss, and bacterial and chemical toxins would increase the number of white blood cells, especially neutrophils, particularly in acute infections (29). Furthermore, haematological findings of this study were within normal limits in all of the groups during the course of the experiment.

Hove et al. (19) demonstrated that TSCFAs in faeces of a man taking antibiotics were below 10 mmol/L. In addition, Hoverstad et al. (20) reported that after application of clindamycin in man, TSCFA values declined from 62.9 to 7.3 mmol/L during the treatment, after using ampicillin from 62.4 to 47.8 mmol/L, there were no significant differences between groups given metronidazol. Similarly, Rao et al. (32) found that TSCFA values decreased in humans given vancomycin and bacitracin. Furthermore, Gustafsson et al. (17) found that TSCFA values were 14.46 mmol/L in patients positive for Cl. difficile toxin and with diarrhoea due to antibiotic usage, and 9.15 mmol/L in patients with diarrhoea due to antibiotic usage, but negative for Cl. difficile toxin. They also found TSCFA values to be much lower in patients with diarrhoea due to antibiotic usage than in healthy individuals. In accordance with the findings of a previous reports (17, 19), TSCFA values of groups I and II on the day of the onset of diarrhoea were low, as compared with their baseline (P<0.001).

Wisker *et al.* (41) found out that succinate and lactate may be converted into TSCFAs in the colon in cases of diarrhoea in pigs caused by antibiotics. Sucrose, maltase, and lactase disaccharide activities were found to be increased in 7 healthy men given lyophilised *S. boulardii* (8) and sucrose and maltase activities in rats given 100 mg/d of *S. boulardii* (10). Zaouche *et al.* (42)

claimed that significant increases occurred in enzymatic of sucrose, glucoamilase, and aminopeptidase in rats given S. boulardii. Yeast cultures have an appetising effect with their attractive natural taste. They contain vitamin B and an unknown growth factor as well as separate vitamins from the chelate form making them ready for the absorption, secreting such digestive enzymes as protease, lipase, proteinase invertase, cellulase, initiating growth of cellulotic bacteria, and synthesising acetate, which is a presubstance for TSCFAs in fermentation (1, 13). The efficacy of S. boulardii in dogs with lincomycin induced diarrhoea in this study, may be explained by the abovementioned reports, as seen in humans. It has been established that oral use of S. boulardii in humans and rats caused a notable increase in both specific and total activities of enzymes without causing any changes in intestinal mucosal membrane, and these enzymes enable breaking indigestible carbohydrates into TSCFAs, making them absorbable.(22, 35, 37). In the presented study, the TSCFA values in group I were found to be rather low (P<0.001), while values in group II given S. boulardii reached the values of its baseline 1 week after the treatment (P=0.127). When the TSCFA values in dogs of groups I and II were compared, it was found out that there were no significant differences (P=0.336) of the baseline and on the day of the onset of diarrhoea (P=0.878). This data shows that the induction of diarrhoea fits well between groups. It was also established that there were significant differences between the day of diarrhoea cessation (P<0.05) and one week after treatment (P<0.05), which demonstrate the efficacy of the probiotic used in the study.

Rao et al. (32) and Hove et al. (19) mentioned that TSCFA values, which decreased as a result of the deterioration of carbohydrate metabolism in diarrhoea, caused by antibiotics, and would reach its normal level in 1-5 weeks. In this study, it was established in group I, one week after the diarrhoea ceased that TSCFA values were still low compared with the baseline, while they reached their baseline in the second week. S. boulardii as a yeast is said to be resistant to antibiotics, which makes it different from other probiotics of a bacterial origin. It is also said not to be affected by gastric acid and bile secretion and does not change its efficiency and activities (12, 31, 37). In this research, TSCFA values in group III, given S. boulardii and lincomycin together, were found to be decreased insignificantly (P=0.1) from the baseline (157.5 \pm 14.6 mmol/L) to the 10th d (138 \pm 8.1 mmol/L). Since the TSCFA values in group III did not decrease significantly enough to induce diarrhoea, S. boulardii could not accomplish reducing the TSCFA values for preventing diarrhoea.

In conclusion, as it was demonstrated, a decrease in TSCFA content causes diarrhoea in the dog treated with lincomycin. Additionally, *S. boulardii* as probiotic was an effective cure for diarrhoea and to prevent its occurrence when given together with lincomycin.

References

- Atherton D., Robbins S.: Probiotics, A European Perspective. In: Biotechnology in the feed Industry. Ed. T.P. Lyons. Alltech Technical Publication, Kentucky 1987, pp. 167-176.
- Baverud V., Gustafsson A., Franklin A., Lindholm A., Gunnarsson A.: Clostridium difficile associated with acute colitis in mature horses treated with antibiotics. Eq Vet J 1997, 29, 279-284.
- 3. Beniwal R.S., Arena V.C., Thomas L., Narla S., Imperiale T.F., Chaudrey R.A., Ahmad U.A.: A randomized trial of yogurt for prevention of antibiotic-associated diarrhea. Dig Dis Sci 2003, **48**, 2077-2082.
- Berezin E.B.: Treatment and prevention of antibiotic associated diarrhea. Int J Antimicrob Agents 2000, 16, 521-526.
- Boddy V.A., Elmer, W.G., Mcfarland V.L., Levy H.R.: Influence of antibiotics on the recovery and kinetics of Saccharomyces boulardii in rats. Pharm Res 1991, 8, 796-800
- Boon R.J., Beale A.S.: Studies with temocillin in the hamster model of antibiotic-associated colitis. Drugs 1985, 29, 57-63.
- 7. Burrows C.F.: Diseases of the colon, rectum, and anus in the dog and cat. In: Veterinary Gastroenterology. Anderson N.V (ed). Lea & Febiger, Philadelphia 1980, pp. 553-592.
- 8. Buts J.P., Bernasconi P., Craynest M.P.V., Maldague P., Meyer R.D.: Response of human and rat small intestinal mucosa to oral administration of Saccharomyces boulardii. Pediatric Res 1986, **20**, 192-196.
- 9. Buts J.P., De Keyser N., Kolanowski J., Sokol E., Van Hoof F.: Maturation of villus and crypt cell functions in rat small intestine: role of dietary polyamines. Dig Dis Sci 1993, **38**, 1091-1098.
- Buts J.P., Keyser N.D., Raedemaeker L.D.: Saccharomyces boulardii enhances rat intestinal enzyme expression by endoluminal release of polyamines. Pediatric Res 1994, 36, 1-7.
- 11. Castagliuolo L., Papadimitra G., Juffer A., Kelley C.P., LaMont J.T., Pothoulakis C.: Saccharomyces boulardii secretes a protease which inhibits Clostridium difficile toxin A receptor binding and enterotoxicity. Gastroenterology 1993, 104, A678.
- Crezucka, D., Rampal, P.: Experimental effects of Saccharomyces boulardii on diarrheal pathogens. Microbes Infect 2002, 4, 733-739.
- Dawson, K.A.: The use of yeast culture in animals feeds: a scientific application of direct fed microbials and challenges of the future. In: Biotechnology in the Feed Industry, 9th Annual Symposium, Proc Alltech's 1993, pp. 169-171.
- Dumankar A.: General side effects of antibiotics. In: Symposium of practical antibiotic usage. Istanbul, 1997, pp. 73-79.
- Filho-Lima J.V.M., Vieira E.C., Nicoli J.R.: Antagonistic effect of Lactobacillus acidophilus, Saccharomyces boulardii and Escherichia coli combinations against experimental infections with Shigella flexineri and Salmonella enteritidis subsp. Typhimurium in gnotobiotic mice. J Appl Microbiol 2000, 88, 365-370.
- Guilford W.G., Center S.A., Strombeck D.R., Williams D.A., Meyer D.J.: Integration of gastrointestinal functions. In: Strombeck's Small Animal Gastroenterology.
 Ed, W.B. Saunders Company, Philadelphia, 1995, pp.16-17.

- Gustafsson A., Lund-Tonnesen S., Berstad A., Midtvedt T. Norin E.: Faecal short-chain fatty acids in patients with antibiotic-associated diarrhea, before and after faecal enema treatment. Scand J Gastroenterol 1998, 33, 721-727.
- Hines R.S., Dickerson S.: Pseudomembranous enteritis associated with ciprofloxacin and Clostridium difficile in a penguin (Eudyptes chrysolophus). J Zoo Wildlife Med 1993, 24, 553-556.
- Hove H., Tvede M., Mortensen B.: Antibiotic-associated diarrhea, Clostridium difficile, and short-chain fatty acids. Scand J Gastroenterol 1996, 31, 688-693.
- Hoverstad T., Carlstedt-Duke B., Lingaas E., Midtvedt T., Norin K.E., Saxerholt H., Steinbakk M.: Influence of ampicillin, clindamycin, and metronidazole on faecal excretion of short-chain fatty acids in healthy subjects. Scand J Gastroenterol 1986, 21, 621-626.
- Hoverstad T., Fausa O., Bjorneklett A., Bohmer T.: Shortchain fatty acids in normal human faeces. Scand J Gastroenterol 1984, 19, 375-381.
- Högenaur C., Hammer H.F., Krejs G.J., Reisinger E.C.: Mechanisms and management of antibiotic-associated diarrhea. Clin Infec Dis 1998, 27, 702-710.
- Hurley B.W., Nyugen C.C.: The spectrum of pseudomembranous enterocolitis and antibioticassociated diarrhea. Arch Intern Med 2002, 162, 2177-2184
- Jabbar A., Wright R. A.: Gastroenteritis and antibioticassociated diarrhea. Prim Care Clin Office Pract 2003, 30, 63-80.
- Koneman E.W., Allen S.B., Janda W.M., Schreckenber P.C., Win W.C.: The Enterobacteriacae. In: Color Atlas and Textbook of Diagnostic Microbiology. 5 Ed, Lippincott, N.Y., 1997, pp. 171-252.
- Marks S.L., Kather E.J.: Bacterial-associated diarrhea in the dog: a critical appraisal. Vet Clin Small Anim 2003, 33, 1029-1060.
- Merrigan M., Sambol S., Johnson S., Gerding D.L.: Susceptibility of hamsters to human pathogenic Clostridium difficile strain B1 following clindamycin, ampicillin or ceftriaxone administration. Anaerob 2003, 9, 91-95.
- Muller E.L., Pitt H.A., George W.L.: Prairie dog model for antimicrobial agent-induced Clostridium difficile diarrhea. Infect Immun 1987, 55, 198-200.
- Noyan A.: Blood Physiology, Yasamda ve Hekimlikte Fizyoloji, Meteksan, 8. Ed. Anonim Co., Ankara 1993, pp. 659-771.
- Orchard J.L., Fekety R., Smith, J.R.: Antibiotic-associated colitis due to Clostridium difficile in a Kodiak bear. Am J Vet Res 1983, 44, 1547-1548.

- Pathoulakis C., Kelly C.P., Joshi M.A., Gao N., Okeane C.J., Castagliuolo L., Lamont J.T.: Saccharomyces boulardii inhibits Clostridium difficile toxin A binding and enterotoxicity in rat ileum. Gastroenterology 1993, 104, 1108-1115.
- 32. Rao S.S.C., Edwards C.A., Austen C.J., Bruce C.: Impaired colonic fermentation of carbohydrate after ampicillin. Gastroenterology 1988, **94**, 928-932.
- Regh M.F., Lu Y.S.: Clostridium difficile in a rabbit following antibiotic therapy for pasteurellosis. J Am Vet Med Assoc 1981, 179, 1296-1297.
- 34. Riley T.V., Adams J.E., Oneill G.L., Bowman R.A.: Gastrointestinal carriage of Clostridium difficile in cats and dogs attending veterinary clinics. Epidemiol Infect 1991, **107**, 659-665.
- Rodriguez A.C.P., Cara D.C., Fretez S.H.G.G., Cunha F.Q., Vieira E.C., Nicoli J.R., Vieira L.Q.: Saccharomyces boulardii stimulates sIgA production and the phagocytic system of gnotobiotic mice. J Appl Microbiol 2000, 89, 404-414.
- Rothman S.W.: Presence of Clostridium difficile toxin in guinea pigs with penicillin-associated colitis. Med Microbiol Immunol (Berl.) 1981, 169, 187-196.
- 37. Surawicz C.M., Elmer G.W., Speelman P., Mcfarland L.V., Chinn J., Belle G.V.: Prevention of antibiotic-associated diarrhoea by Saccharomyces boulardii: a prospective study. Gastroenterology 1989, **96**, 981-988.
- Tsukahara T., Iwasaki Y., Nakayama K., Ushidi K.: Microscopic structure of the large intestinal mucosa in piglets during an antibiotic-associated diarrhea. Vet Med Sci 2003, 65, 301-306.
- Twedt D.C.: Clostridium perfiringens-associated diarrhoea in dogs. In: Proc Am Col Vet Int Med Forum, Washington DC, 1993, pp. 121-125.
- Weber A., Kroth P., Heil G.: Occurrence of Clostridium difficile in faeces of dogs and cats. J Vet Med Series (B), 1989, 36, 568-576.
- Wisker E., Daniel M., Rave G., Feldheim W.: Short-chain fatty acids produced in vitro from fibre residues obtained from mixed diets containing different breads and in human faeces during the ingestion of the diets. Br J Nutr 2000, 84, 31-37.
- Zaouche A., Loukil C., Lagausie P.D., Macry J., Fitoussi F., Bernasconi P., Bingen E., Cezard J.P.: Effects of oral Saccharomyces boulardii on bacterial overgrowth, translocation, and intestinal adaptation after small-bowel resection in rats. Scand J Gastroenterol 2000, 35, 160-165.